

First Experimental Demonstration of the Multipotential Carcinogenic Effects of Aspartame Administered in the Feed to Sprague-Dawley Rats

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The National Institute of Environmental Health Sciences National Institutes of Health U.S. Department of Health and Human Services First Experimental Demonstration of the Multipotential Carcinogenic Effects of Aspartame Administered in the Feed to Sprague-Dawley Rats

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## Abbreviations:

ADI:	Acceptable Daily Intake
APM:	Aspartame
CMCRC/ERF:	Cesare Maltoni Cancer Research Center/European Ramazzini
	Foundation
DKP:	Diketopiperazine
FDA:	Food and Drug Administration
HE:	Hematoxylin –Eosin
IARC	International Agency for Research on Cancer
MSA:	Monosodium Aspartate
MTBE	Methyl- <i>tert</i> -butyl ether
pmm	Post mortem modifications

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#### ABSTRACT

The Cesare Maltoni Cancer Research Center of the European Ramazzini Foundation has conducted a long-term bioassay on aspartame (APM), a widely used artificial sweetener. APM was administered with feed to 8 week-old Sprague-Dawley rats (100-150/sex/group), at concentrations of 100,000; 50,000; 10,000; 2,000; 400; 80 or 0 ppm. The treatment lasted until natural death, at which time all deceased animals underwent complete necropsy. Histopathological evaluation of all pathological lesions and of all organs and tissues collected was routinely performed on each animal of all experimental groups.

The results of the study show for the first time that APM, in our experimental conditions, causes: 1) an increased incidence of malignant tumor-bearing animals with a positive significant trend in males ( $p\leq0.05$ ) and in females ( $p\leq0.01$ ), in particular those females treated at 50,000 ppm ( $p\leq0.01$ ); 2) an increase in lymphomas and leukemias with a positive significant trend in both males ( $p\leq0.05$ ) and females ( $p\leq0.01$ ), in particular in females treated at doses of 100,000 ( $p\leq0.01$ ), 50,000 ( $p\leq0.01$ ), 10,000 ( $p\leq0.05$ ), 2,000 ( $p\leq0.05$ ), 400 ( $p\leq0.01$ ) ppm; 3) a statistically significant increased incidence, with a positive significant trend ( $p\leq0.01$ ) of transitional cell carcinomas of the renal pelvis and ureter and their precursors (dysplasias) in females treated at 100,000 ( $p\leq0.01$ ), 50,000 ( $p\leq0.01$ ), 10,000 ( $p\leq0.01$ ), 2,000 ( $p\leq0.05$ ) and 400 ppm ( $p\leq0.05$ ); and 4) an increased incidence of malignant schwannomas of peripheral nerves with a positive trend ( $p\leq0.05$ ) in males.

The results of this mega-experiment indicate that APM is a multipotential carcinogenic agent, even at a daily dose of 20 mg/kg b.w., much less than the current acceptable daily intake (ADI). On the basis of these results, a re-evaluation of the present guidelines on the use and consumption of APM is urgent and cannot be delayed.

#### INTRODUCTION

Nowadays consumers are increasingly concerned about the quality and safety of many products present in the diet of industrialized countries, in particular the use of artificial sweeteners, flavorings, colorings, preservatives and dietary supplements. General apprehension also exists regarding the possible long-term health effects of the raw materials and technologies used for the packaging, sterilization and distribution of foods. Of particular concern are the potential carcinogenic effects of these products and processes.

The experimental and epidemiological data currently available to evaluate the above carcinogenic risks are insufficient and often unreliable, due to the inadequate planning and conduct of previous experiments. This inadequacy, combined with the general limited knowledge about the safety/potential carcinogenic effects of substances widely present in the industrialized diet, motivated the design of an integrated project of mega-experiments in 1985 at the Cesare Maltoni Cancer Research Center (CMCRC) of the European Ramazzini Foundation (ERF). The products studied are reported in Table 1. Products and agents selected for the project were those for which committee debate and opinions often acted as surrogates for good laboratory work. Over the course of the project, up to now, 32 long-term bioassays have been performed using over 25,000 rodents. Studies have evaluated the carcinogenicity of 12 different products, including the artificial sweetener Aspartame (APM).

The following report presents the results of the mega-experiment on the carcinogenicity of APM in which the sweetener was administered in feed to Sprague-Dawley rats for the life span. APM, the methyl ester of the dipeptide L- $\alpha$ -aspartyl-L-phenylalanine (C<sub>14</sub>H<sub>18</sub>N<sub>2</sub>O<sub>5</sub>), is a widely used artificial sweetener with a molecular weight of 294.3. Under particular conditions (extreme pH, high temperature, lengthy storage times) APM may be contaminated by the diketopiperazine cycloaspartylphenylalanine (DKP) (Butchko et al. 2002a).

For more than 30 years, APM has been widely used as a food additive due to its very strong, sweet taste. The sweetening power of APM is estimated to be 200 times that of sucrose, whereas saccharin and cyclamate are 300 and 30 times sweeter, respectively (Mazur 1984).

Initial commercial approval of APM in the United States was granted by the FDA in 1974. The FDA later approved the limited use of APM in solid foods in 1981 and extended this authorization to soft drinks in 1983. APM was eventually approved as a general sweetener in 1996 (FDA 1981, 1983, 1996). In the European Union, the safe use of APM was authorized in 1994.

After saccharin, APM is the second most used artificial sweetener in the world. It is estimated that more than 8,000 tons of APM are consumed each year in the USA (Hazardous Substances Data Bank 2005). In terms of world consumption, APM represents 62% of the value of the intense sweetener market (Fry 1999).

APM is found in more than 6,000 products, including carbonated and powdered soft drinks, hot chocolate, chewing gum, candy, desserts, yoghurt, table-top sweeteners and in some pharmaceutical products, such as vitamins and sugar-free cough drops, and is estimated by the Aspartame Information Center (2005) to be consumed by over 200 million people worldwide.

The average APM daily intake in the general population has been shown, by dietary surveys performed in the United States among APM consumers during the period 1984-1992, to range from 2 to 3 mg/kg of body weight. Consumption by 2 to 5 year-old children and by females of childbearing age in these surveys ranged from about 2.5 to 5 mg/kg b.w./day (Butchko et al. 2002b). APM intake was also monitored in several other regions, including 7 European countries. Although survey methodologies may have differed, the APM intake was remarkably consistent across studies and was well below the Acceptable Daily Intake (ADI) both in the United States (50 mg/kg b.w.) and in Europe (40 mg/kg b.w.) (Butchko et al. 2002b).

Investigations into the metabolism of APM have shown that, in rodents, non-human primates and humans, it is metabolized in the gastrointestinal tract into three constituents: aspartic acid, phenylalanine and methanol, which are absorbed into the systemic circulation (Ranney et al. 1976). For each molecule of APM, one molecule of each constituent is produced. After absorption, they are then utilized, metabolized and/or excreted by the body following the same metabolic pathways as when consumed through the ordinary diet, namely: aspartate is transformed into alanine plus oxaloacetate (Stegink 1984); phenylalanine is transformed mainly into tyrosine and, to a smaller extent, phenylethylamine and phenylpyruvate (Harper 1984); while methanol is transformed into formic acid (Opperman 1984).

It has been reported that APM is not genotoxic in the following tests: dominant lethal mutation assay in rats, host-mediated assay in rats and mice, *in vivo* cytogenetic assay in rats, the Ames test (Kotsonis and Hjelle 1996). Results of an assay to measure induction of unscheduled DNA synthesis in rat hepatocytes treated with APM *in vitro* were negative,

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indicating the absence of APM-induced DNA damage (Jeffrey and Williams 2000). *In vivo*, a mixture of aspartame (up to 350 mg/kg) and a second sweetener, acesulfame potassium (up to 150 mg/kg), was reported to be negative in a test for the induction of chromosomal aberration in bone marrow cells of male Swiss mice, when administered by gavage. However a dose-related increase in the percentage of cells with chromosomal aberrations was noted with increasing doses of the two sweeteners, but the increase was not statistically significant (Mukhopadhyay et al. 2000).

Two long-term feeding carcinogenicity bioassays on APM were performed on rats and one on mice in the early 1970s by the producer Searle & Co. Results were reviewed by the FDA and then summarized in the Federal Register of 1981 (FDA 1981). To date, the details of the experiments have not been published.

In the first study, groups of 40 male and 40 female Sprague-Dawley rats were treated with 1, 2, 4, or 6 to 8 g/kg b.w./day of APM in the diet. The treatment started at 4 weeks of age and lasted for a period of 104 weeks. A control group of 60 rats per sex was fed the same diet without APM. At the end of the treatment, all surviving animals were sacrificed and their brains, as well as other organs (not specified in the report), were examined histologically. Brain tumors were observed in 7/155 (4.5%) exposed males vs 1/59 (1.7%) controls, and in 5/158 (3.2%) exposed females vs 0/59 (0%) in controls. Overall, the FDA considered the study to be negative with regard to the carcinogenicity of APM.

In the second study, groups of 40 male and 40 female Sprague-Dawley rats were exposed to APM, at doses of 2 and 4 g/kg b.w./day, through their mothers' diet both *in utero* and during lactation, and then for 104 weeks with APM in their own diets. A control group of 60 rats per sex was fed the same diet without APM. The animals were necropsied

at the time of death or at 104 weeks after weaning. Three brain tumors were observed among control males and one among control females. Brain tumors were also observed in two males and one female in the 2 g/kg b.w. group, and in one male and one female in the 4 g/kg b.w. group. Again, the FDA considered the study to be negative with regard to the carcinogenicity of APM.

Regarding the third chronic APM study, in this case performed on mice, the FDA reported that the results did not show any treatment-related carcinogenic effect. In this experiment, as reported by Molinary (1984), groups of 36 male and 36 female mice were fed 1, 2, 4 g/kg b.w./day until 110 weeks of age. A group of 72 males and 72 females served as the control. There were no treatment-related effects on survival and behavior, nor were any lesions recorded during macroscopic or microscopic analysis.

An APM carcinogenicity study was also conducted in Japan during this period (Ishii 1981; Ishii et al. 1981). Groups of 86 male and 86 female Wistar rats were treated with APM in feed at doses of 0, 1, 2, or 4 g/kg b.w./day from 6 to 110 weeks of age. No increase in the incidence of brain tumors was observed in the treated groups as compared to the controls. Exhaustive experimental details of this study were not published.

Epidemiological studies to evaluate the relationship between APM intake and cancer development in humans are not currently available.

Although all of the aforementioned studies were considered negative with respect to the carcinogenicity of APM, in our opinion, these studies did not comply with today's basic requirements for testing the carcinogenic potential of a physical or chemical agent, in particular concerning the number of animals for each experimental group and the duration of the experiment until 110 weeks of age of the animals.

For these reasons, and in light of the ever increasing diffusion of APM in the diet of industrialized countries (particularly in products consumed by young children and pregnant women), we considered it important to perform a mega-experiment following today's internationally recognized Good Laboratory Practices for carcinogenicity bioassays and, more specifically, the life-span carcinogenicity bioassay design followed for many years at the CMCRC and described in previous publications (Soffritti et al. 1999; Soffritti et al. 2002c).

#### MATERIALS AND METHODS

The APM, as a food grade material, was produced by Nutrasweet and supplied by Giusto Faravelli S.p.A. in Milan, Italy. Its purity was more than 98%: DKP was less than 1.5% and L-phenylalanine was less than 0.5%. The method used to determine APM purity was an infrared absorption spectrophotometer assay. An assumed daily intake by humans of 5,000; 2,500; 500; 100; 20; 4 or 0 mg/kg b.w. was simulated by adding APM to the standard Corticella diet, used for 30 years at the CMCRC/ERF Laboratory, at concentrations of 100,000; 50,000; 10,000; 2,000; 400; 80; or 0 ppm. The APM daily assumption in mg/kg b.w. was calculated considering the average weight of a rat for the duration of the experiment as 400 g, and the average consumption of feed as 20 g per day, both for males and females. APM was administered with feed *ad libitum* to Sprague-Dawley rats (100-150/sex/group). The experiment started when the animals were 8 weeks-old. The treatment lasted until natural death. Control animals received the same feed without APM. The experiment was conducted according to the Italian law regulating the

use of animals for scientific purposes (Decreto Legislativo 116 1992), which provides the guidelines on how to treat animals humanely and without suffering.

Rodents used for the experiment were male (M) and female (F) Sprague-Dawley rats from the colony of the CMCRC/ERF. This colony of rats has been employed for various experiments in the laboratory for nearly 30 years and extensive historical data are available on the tumor incidence among untreated rats. All control animals were monitored for feed and water consumption and body weight for their life span and, upon death, underwent complete necropsy and histopathological evaluation.

The health status of the animals was regularly checked by the veterinarians of the Local and National Health Services. Before matching, the breeders were clinically observed for their health status, in order to exclude any diseased animals and the experimental animals were clinically examined monthly until the end of the experiment.

At 4-5 weeks of age, after weaning, the experimental animals were randomised in order to have no more than one male and one female from each litter in the same group. They were then housed, in groups of 5, in makrolon cages (41x25x15 cm), with stainless-steel wire tops and a shallow layer of white wood-shavings as bedding, and kept in rooms destined only to this experiment, at a temperature of  $23 \pm 2^{\circ}$ C and relative humidity of 50-60%, respectively.

Once a week for the first 13 weeks, then every two weeks until 110 weeks of age, the mean daily drinking water and feed consumption were measured per cage, and body weight measured individually. Measurement of body weight continued every 8 weeks until the end of the experiment. The animals were clinically examined for gross changes every 2 weeks for the entire duration of the experiment. In order to evaluate the status and behaviour of

the animals and to limit the *post mortem* modifications (pmm), a patrol was performed three times daily from Monday to Friday and twice on Saturdays and Sundays and festivities. Dead animals were registered and kept refrigerated at 4°C until necropsy. Based on this procedure (part of our longstanding Standard Operating Procedures, SOP) very few animals were affected by pmm and, on very rare occasions, this interfered with the ability to histologically diagnose and interpret some lesions.

The biophase ended at 151 weeks, with the death of the last animal at the age of 159 weeks. Upon death, the animals underwent complete necropsy. Histopathology was routinely performed on the following organs and tissues of each animal from each group: skin and subcutaneous tissue, mammary gland, the brain (3 sagittal sections), pituitary gland, Zymbal glands, salivary glands, Harderian glands, cranium (five sections, with oral and nasal cavities and external and internal ear ducts), tongue, thyroid, parathyroid, pharynx, larynx, thymus and mediastinal lymph nodes, trachea, lung and mainstem bronchi, heart, diaphragm, liver, spleen, pancreas, kidneys, adrenal glands, oesophagus, stomach (fore and glandular), intestine (four levels), urinary bladder, prostate, gonads, interscapular brown fat pad, subcutaneous and mesenteric lymph nodes, and other organs or tissues with pathological lesions. All organs and tissues were preserved in 70% ethyl alcohol, except for bones which were fixed in 10% formalin and then decalcified with 10% formaldehyde and 20% formic acid in water solution. The normal specimens were trimmed, following the CMCRC/ERF Laboratory SOP. Trimmed specimens were processed as paraffin blocks, and 3-5 micron sections of every specimen were obtained.

Sections were routinely stained with Hematoxylin-Eosin (HE). Immunohistochemical staining for S100 was performed to characterize malignant Schwannoma, while

chromogranin A staining was used to characterize olfactory neuroblastoma. For S100 staining, a polyclonal rabbit Anti-S100 (Dakocytomation code no. Z0311) was used as primary antibody while, for chromogranin A staining, a polyclonal rabbit anti-human chromogranin A (Dakocytomation code no. N1535) was used (Information Centre for Immunohistochemistry 2005).

Two statistical tests were used to analyze neoplasm and non neoplastic lesion incidence data. The Cochran-Armitage trend test (Armitage 1971; Gart et al. 1979) was used to test for linear trends in tumor incidence. Also used was the poly K-test (Bailer and Portier 1988; Portier and Bailer 1989; Piegorisch and Bailer 1997), a survival-adjusted quantal response modification of the Cochran-Armitage test that takes survival into account. The test used and the p-values are reported in the tables.

### RESULTS

The study proceeded smoothly without unexpected occurrences. No differences were observed in water consumption between the treated and the untreated groups, whereas a dose-related difference in feed consumption was observed between the various treated groups and the control group in both males and females (Figure 1A, B). No substantial differences in mean body weight were observed between the treated and control groups, apart from a slight decrease in females treated at 100,000 ppm (Figure 1C). No substantial difference in survival was observed among the groups (Figure 1D, E).

No evident behavioral changes were observed among treated animals compared to the controls. In animals exposed to the highest dose of APM, yellowing of the coat was

observed: this change had previously been observed in our laboratory in rats exposed to formaldehyde administered with drinking water (Soffritti et al. 2002b).

The carcinogenic effects of APM are reported in Table 2 for males and Table 3 for females. Multiple tumors of different type and site, of different type in the same site, of the same type in bilateral organs, of the same type in the skin, in the subcutaneous tissue, and in mammary glands, or at distant sites of diffuse tissue (i.e. bones and skeletal muscle) were plotted as single/independent tumors. Multiple tumors of the same type in the same tissue and organ, apart those above mentioned, were plotted only once.

<u>Total malignant tumors.</u> The incidence of malignant tumor-bearing animals occurred with a significant positive trend in males ( $p \le 0.05$ ) and in females ( $p \le 0.01$ ) as reported in Tables 2 and 3. A statistically significant increase of the incidence of malignant tumors was observed in females treated at 50,000 ppm ( $p \le 0.01$ ) compared to the control group (Table 3). Tumor types which contributed most are presented as follows.

<u>Lymphomas-leukemias.</u> The data on the occurrence of lymphomas-leukemias, reported in Tables 2 and 3, indicate that APM causes a significant positive trend in males ( $p \le 0.05$ ) and in females ( $p \le 0.01$ ). When compared to untreated control groups, the increased incidence of lymphomas-leukemias in treated females was statistically significant at doses of 100,000 ( $p \le 0.01$ ), 50,000 ( $p \le 0.01$ ), 10,000 ( $p \le 0.05$ ), 2,000 ( $p \le 0.05$ ) and 400 ( $p \le 0.01$ ) ppm. The most frequent histocytotypes observed in the experiment were lymphoimmunoblastic lymphomas, mainly involving lung and mediastinal/peripheral nodes, and histiocytic sarcomas, involving mainly lung, liver, spleen and nodes. The distribution of lymphomas-leukemias by histocytotypes is presented in Table 4. The differential diagnoses were based on the morphological criteria followed in our laboratory for several decades and are in line with the guidelines of the International Classification of Rodent Tumors (IARC 1993). Lymphomas-leukemias (this term includes all types of hemolymphosarcomas and leukemias) are neoplasias arising from hemolymphoreticular tissues, and their aggregation is widely used in experimental carcinogenesis. The reason is, as has been stated, that both solid and circulating phases are present in many lymphoid neoplasms, and distinction between them is artificial (Harris et al. 2001).

Preneoplastic and neoplastic lesions of the renal pelvis and ureter. The incidences of preneoplastic and neoplastic lesions of the transitional cell epithelium of the renal pelvis and ureter are reported in Tables 2 and 3. A dose-related increase in the incidence of dysplastic hyperplasias and dysplastic papillomas of the renal pelvis and ureter were observed in females. Carcinomas in females occurred with a positive trend ( $p \le 0.05$ ) and the incidence in females exposed at 100,000 ppm was significantly higher (p≤0.05) compared to the controls. Carcinomas were also observed among males treated at 100,000, 50,000, 10,000, and 2,000 ppm. In females, dysplastic lesions and carcinomas combined show a significant positive trend ( $p \le 0.01$ ) and a statistically significant increase in those treated at 100,000 (p≤0.01), 50,000 (p≤0.01), 10,000 (p≤0.01), 2,000 (p≤0.05) and 400 ppm (p≤0.05). A threefold increase is also observed in the 80 ppm treated group. We did not observe substantial differences in the incidence of inflammation between males and females treated at the different doses, as compared to controls. Increased incidence of calcification was observed in females, particularly in those treated at 100,000 ppm (39%), 50,000 ppm (25%) and 10,000 ppm (19%), when compared to controls (8%); this effect was not observed in males. It should be pointed out that, while transitional cell carcinomas of the renal pelvis and ureter are extremely rare in male and female untreated rats, the APM male and female

groups had a total of 21 transitional cell carcinomas of the renal pelvis, whereas the controls had none. Microscopically the carcinomas were invading, with various levels of extension, the papilla and the kidney parenchyma; the cells were of transitional type and several mitotic figures were present (Figure 2A, B).

*Malignant Schwannomas of peripheral nerves.* As shown in Table 2, the incidence of malignant schwannomas of the peripheral nerves occurred with a positive trend ( $p \le 0.05$ ) in males. In females, 9 malignancies were observed among treated animals of the different dosage groups and none among the controls (Table 3). All lesions, in males and females, diagnosed as malignant schwannomas were positive for S100 staining. The most frequent site of origin of the malignant schwannomas was in the cranial nerves (72%); the other cases arose at the spinal nerve roots. Microscopically, malignant schwannomas were observed in 3 males treated at the highest dose. The metastases were found in submandibular lymph nodes in 2 cases, and in 1 case, the tumor metastatized to the lung and to the liver. Histologically the feature of malignant schwannomas was Antoni B type (Figure 2C, D).

<u>Preneoplastic and neoplastic lesions of the olfactory epithelium.</u> Incidence of hyperplasia of the olfactory epithelium increased with a significant positive trend in males and females. The observed incidences were respectively: 14.0% and 18.0% in males and females exposed at 100,000 ppm; 12.0% and 21.0% at 50,000 ppm; 7.0% and 17.0% at 10,000 ppm; 2.7% and 8.7% at 2,000 ppm; 6.0% and 7.3% at 400 ppm; 2.0% and 3.3% at 80 ppm; and 0.7% and 4.0% at 0 ppm. The differences were statistically significant ( $p\leq0.01$ ) at 100,000, 50,000 and 10,000 ppm in both males and females and also, in males, at 400 ppm. It is noteworthy that among females treated at the highest dose, one case of

dysplastic hyperplasia, one adenoma and one olfactory neuroblastoma were observed. The neuroblastoma invaded the cranium, compressing the forebrain and was positive for chromogranin A immunohistochemical staining.

*Brain malignant tumors.* Concerning the incidence of brain malignant tumors, it should be noted that, as previously reported (Soffritti et al. 2005), 12 malignant tumors (10 gliomas, 1 medulloblastoma and 1 meningioma) were observed, without dose relationship, in male and female APM-treated groups, while none were observed in controls.

<u>Other malignant tumors.</u> The other malignant tumors were among those commonly observed in Sprague-Dawley rats, apart from 2 transitional cell carcinomas of the bladder observed in males exposed to 10,000 ppm and 1 in females exposed to 2,000 ppm and none among the controls. Since this type of tumor is extremely rare among the historical controls of our colony of Sprague-Dawley rats, this occurrence cannot be disregarded.

*Historical controls.* Over the last 20 years, in our laboratory, when we consider only groups of 100 or more animals per sex, the numbers of the untreated males and females total 1934 and 1945 respectively. Concerning the renal pelvis and ureter transitional cell carcinomas, no carcinomas were observed in either males or females. The overall incidence of malignant schwannomas was 0.5% (0-2.0%) in males and 0.1% (0-1.0%) in females. The overall incidence of lymphomas-leukemias was 20.7% (8.0-30.9%) in males and 12.4% (7.0-18.4%) in females. The overall incidence of olfactory neuroblastoma was 0.1% (0-1.8%) in both males and females.

When we also consider control groups of less than 100 animals per sex, the numbers of untreated males and females total 2265 and 2274 respectively. The overall incidence of the renal pelvis and ureter transitional cell carcinomas was 0.04% (0-1.0%) in females, while

no carcinomas were observed in males. The overall incidence of malignant schwannomas was 0.4% (0-2.0%) in males and 0.1% (0-2.0%) in females. The overall incidence of lymphomas-leukemias was 20.6% (8.0-30.9%) in males and 13.3% (4.0-25.0%) in females. The overall incidence of olfactory neuroblastomas was 0.1% (0-1.8%) in both males and females.

#### DISCUSSION

The mega-experiment performed in our laboratory on APM (administered with feed to Sprague-Dawley rats from 8 weeks-old until natural death) has shown for the first time the multipotential carcinogenic effects of this compound. In fact, the results indicate that APM causes, in our experimental conditions: 1) an increased incidence of malignant tumor-bearing animals with a positive significant trend in males ( $p\leq0.05$ ) and in females ( $p\leq0.01$ ), particularly in the females treated at 50,000 ppm ( $p\leq0.01$ ); 2) a statistically significant dose-related increase of the incidence of lymphomas-leukemias in females treated at the doses of 100,000 ( $p\leq0.01$ ), 50,000 ( $p\leq0.01$ ), 10,000 ( $p\leq0.05$ ), 2,000 ( $p\leq0.05$ ) and 400 ( $p\leq0.01$ ) ppm and a positive significant trend in both males ( $p\leq0.05$ ) and females ( $p\leq0.01$ ); 3) in females, dysplastic lesions and carcinomas of the renal pelvis and ureter combined show a significant positive trend ( $p\leq0.01$ ), 10,000 ( $p\leq0.01$ ), 2,000 ( $p\leq0.05$ ) and 400 ( $p\leq0.05$ ) ppm; and 4) an increased incidence of malignant schwannomas of the peripheral nerves with a positive trend ( $p\leq0.05$ ) in males.

The increase in lymphomas-leukemias in APM-treated females could be related to its metabolite methanol, which is in turn metabolized to formaldehyde in both humans and rats

(Ranney et al. 1976). In fact, previous experiments performed at the CMCRC Laboratory have shown that: 1) methanol administered in drinking water, at doses ranging from 20,000 to 500 ppm, induced a statistically significant increase in the incidence of lymphomas-leukemias in female rats, (Soffritti et al. 2002a); 2) a dose-related increase in the incidence of lymphomas-leukemias was also observed in females treated with formaldehyde, administered in drinking water at doses ranging from 1,500 to 50 ppm (Soffritti et al. 1989; Soffritti et al. 2002b); and 3) the same effect was observed in females treated with the gasoline oxygenated additive methyl-*tert*-butyl-ether (MTBE), which metabolizes to methanol (Belpoggi et al. 1995).

The important role of formaldehyde in the induction of hematological malignancies in rodents is further highlighted by these results. In a recent re-evaluation of the carcinogenicity of formaldehyde by the International Agency for Research on Cancer (IARC), strong (although not considered sufficient) evidence of an association between formaldehyde exposure and leukemias in humans was found (IARC, in press).

Moreover, carcinogenic effects for the renal pelvis and ureter, peripheral nerves and proliferative changes of the olfactory epithelium were not observed in the long-term bioassays performed in the same conditions at the CMCRC on methanol, MTBE or formaldehyde. To investigate if the other two metabolites of APM are responsible in inducing these lesions, it is of paramount importance to perform adequate life-span carcinogenicity studies on aspartic acid or phenylalanine.

It is worthy of note that, in a long-term carcinogenicity study on monosodium aspartate (MSA) administered with drinking water to groups of 50 male and 50 female Fischer-344 rats (beginning at 6 weeks of age for 100 weeks and then sacrificed), a dose-related

hyperplasia of the renal pelvis was observed in males and in females (Kitahori et al. 1996). The same effect was found, by the same group of investigators, in another study in which MSA was administered in drinking water to groups of male and female Fischer-344 rats to evaluate its promoting activity of carcinogenesis of the transitional epithelium of the renal pelvis (Kitamura et al. 1996). In both studies, clear evidence was provided of a relationship between MSA treatment and transitional cell hyperplasia. The authors indicated that calcification could have an important role in inducing simple and papillary hyperplasia of the renal pelvis transitional cell epithelium and, consequently, in the induction of transitional cell tumors. In our study, performed on 1,800 Sprague-Dawley rats, which are less susceptible to the spontaneous development of nephropathies than Fischer rats, we observed a dose-related, statistically significant increase in the incidence of dysplastic hyperplasia and carcinoma of the renal pelvis in females, but none in males, when compared to the controls. The fact that we observed an increased incidence of kidney calcification in females and not in males, when compared to the controls, gives added weight to the hypothesis that aspartic acid may cause preneoplastic and neoplastic lesions of the renal pelvis, and that calcification may be the mechanism responsible for this effect.

The carcinogenic effects of APM observed in our experiment are in contrast with the results obtained with long-term carcinogenicity bioassays, performed almost 30 years ago, on Sprague-Dawley rats, which did not reveal APM to have any carcinogenic effects (FDA, 1981). There are several reasons which can explain this difference. First of all, in our experiment the number of animals per sex per group was much greater, allowing a more thorough and reliable statistical analysis. Secondly, in our experiment, rodents were not killed at 110 weeks of age, but rather were observed until natural death, to allow APM to

fully express its carcinogenic potential. Had we stopped the experiments at 110 weeks of age, we would most likely never have demonstrated the carcinogenicity of important industrial compounds such as, xylenes, mancozeb, vinyl acetate monomer (Soffritti et al. 2002c) and toluene (Soffritti 2004).

Finally, concerning the absence of carcinogenic effects observed in the experiment performed on Wistar rats (Ishii 1981; Ishii et al. 1981), it cannot be disregarded that this strain is more resistant than Sprague-Dawley rats to developing cancer, a characteristic shown in our experiments on benzene (Maltoni et al. 1989). Moreover, the aforementioned experiment on Wistar rats was terminated at the age of 110 weeks. Given these differences, the results of the Wistar rat study are not comparable with those performed on Spague-Dawley rats.

#### CONCLUSIONS

Our study has shown that APM is a multipotential carcinogenic compound whose carcinogenic effects are evident even at a daily dose of 20 mg/kg b.w., much less than the current ADI for humans in Europe (40 mg/kg b.w.) and in the United States (50 mg/kg b.w.).

It has been shown that the results of carcinogenicity bioassays in rodents are consistent predictors of human cancer risks (Huff 1999; Tomatis et al. 1989; Rall 1995). The results of our study therefore call for an urgent re-examination of the present guidelines on the use and consumption of APM. The decision to use experimental data to protect public health is important as the time span of widespread aspartame use is still too brief to have produced solid epidemiologic data. Moreover, it is unlikely that sufficient epidemiological data will

be available in the near future, given the difficulty of finding a control group that has not been exposed to this widely diffused compound.

#### REFERENCES

- Armitage P. 1971. Statistical Methods in Medical Research. New York: John Wiley & Sons.
- Aspartame Information Center. 2005. Available: http://www.aspartame.org. [accessed 27 October 2005].
- Bailer AJ, Portier CJ. 1988. Effects of treatment-induced mortality and tumor-induced mortality on tests for carcinogenicity in small samples. Biometrics 44:417-431.
- Belpoggi F, Soffritti M, Maltoni C. 1995. Methyl-tertiary-butyl ether (MTBE), a gasoline additive, causes testicular and lymphohaematopoietic cancers in rats. Toxicol Ind Health 11:119-149.
- Butchko HH, Stargel WW, Comer CP, Mayhew DA, Benninger C, Blackburn GL et al. 2002a. Preclinical safety evaluation of aspartame. Regul Toxicol Pharmacol 35:S7-S12.
- Butchko HH, Stargel WW, Comer CP, Mayhew DA, Benninger C, Blackburn GL et al. 2002b. Intake of aspartame vs the acceptable daily intake. Regul Toxicol Pharmacol 35:S13-S16.
- Decreto Legislativo 116. 1992. Attuazione della direttiva n. 86/609/CEE in materia di protezione degli animali utilizzati a fini sperimentali o ad altri fini scientifici. [In Italian]. Supplemento ordinario alla Gazzetta Ufficiale 40:5-25.
- Food and Drug Administration. 1981. Aspartame: Commissioner's final decision. Fed Regist 46:38285-38308.
- Food and Drug Administration. 1983. Food additives permitted for direct addition to food for human consumption: aspartame. Fed Regist 48:31376-31382.

- Food and Drug Administration. 1996. Food additives permitted for direct addition to food for human consumption; aspartame. Fed Regist 61:33654-33656.
- Fry J. 1999. The world market for intense sweeteners. World Rev Nutr Diet 85:201-211.
- Gart JJ, Chu KC, Tarone RE. 1979. Statistical issues in interpretation of chronic tests for carcinogenicity. J Natl Cancer Inst 62:957-974.
- Harper AE. 1984. Phenylalanine metabolism. In: Aspartame Physiology and Biochemistry (Stegink LD, Filer LJ Jr, eds). New York: Dekker, 77-109.
- Harris NL, Jaffe ES, Vardiman JW, Stein H, Diebold J, Müller-Hermelink HK, *et al.* 2001.WHO Classification of tumors of haematopoietic and lymphoid tissues: Introduction. InTumors of Haematopoietic and Lymphoid Tissues (Jaffe ES, Harris NL, Stein H,Vardiman JW eds). Lyon: IARC Press, 12-13.
- Hazardous Substances Data Bank. 2005. Available: <u>http://toxnet.nlm.nih.gov/</u> [accessed 3 August 2005].
- Huff J. 1999. Long-term chemical carcinogenesis bioassays predict human cancer hazards. Issues, controversies, and uncertainties. In Uncertainty in the Risk Assessment of Environmental and Occupational Hazards, 24-26 September 1998, Bologna, Italy. Ann NY Acad Sci, 895: 56-79.
- IARC. 1993. International classification of rodent tumors. Part I: the rat.4.Haematopoietic system. IARC Scientific Publications N° 122. Lyon, France. International Agency for Research on Cancer.
- IARC. In press. Formaldehyde, 2-Butoxyethanol and 1-tert-Butoxy-2-Propanol. Monogr Eval Carcinog Risks Hum.

- Information Centre for Immunohistochemistry. 2005 Available: <u>http://www.ihcworld.com</u> [accessed 1 April 2005].
- Ishii H.1981. Incidence of brain tumors in rats fed aspartame. Toxicol Lett 7:433-437.
- Ishii H, Koshimizu T, Usami S, Fujimoto T. 1981. Toxicity of aspartame and its diketopiperazine for Wistar rats by dietary administration for 104 weeks. Toxicology 21: 91-94.
- Jeffrey AM, Williams GM. 2000. Lack of DNA-damaging activity of five non-nutritive sweeteners in the rat hepatocyte/DNA repair assay. Food Chem Toxicol 38:335-338.
- Kitahori Y, Kitamura M, Konoshi N, Matsuda H, Tao M, Matsui E et al. 1996.Carcinogenicity study of monosodium aspartate in Fisher 344 rats: 100 weeks treatment.J Toxicol Pathol 9:161-168.
- Kitamura M, Konishi N, Kitahori Y, Fukushima Y, Yoshioka N, Hiasa Y. 1996. Promoting effect of monosodium aspartate, but not glycine, on renal pelvis and urinary bladder carcinogenesis in rat induced by *N*-Butyl-*N*-(4-Hydroxybutyl)nitrosamine. Toxicol Pathol 24:573-579.
- Kotsonis FN, Hjelle JJ. 1996. The safety assessment of aspartame: scientific and regulatory considerations. In: The Clinical Evaluation of a Food Additive: Assessment of Aspartame (Tschanz C, Butchko HH, Stargel WW, Kotsonis FN, eds). Boca Raton: CRC Press, 23-41.
- Maltoni C, Ciliberti A, Cotti G, Conti B, Belpoggi F. 1989. Benzene, an experimental multipotential carcinogen: Results of the long-term bioassays performed at the Bologna Institute of Oncology. Environ Health Perspect 82:109-124.

- Maltoni C, Lefemine G, Belpoggi F, Soffritti M, Lenzi A, Ciliberti A et al. 1997. Risultati di saggi sperimentali di cancerogenicità di acque minerali contenute in bottiglie di PVC, su ratti Sprague-Dawley [in Italian]. Eur J Oncol 6:531-551.
- Mazur RH. 1984. Discovery of aspartame. In: Aspartame Physiology and Biochemistry (Stegink LD, Filer LJ Jr, eds). New York: Dekker, 3-9.
- Molinary SV. 1984. Preclinical studies of aspartame in non primate animals. In: Aspartame Physiology and Biochemistry (Stegink LD, Filer LJ Jr, eds). New York: Dekker, 289-306.
- Mukhopadhyay M, Mukherjee A, Chakrabarti J. 2000. In vivo cytogenetic studies on blends of aspartame and acesulfame-K. Food Chem Toxicol 38:75-77.
- Opperman JA. 1984. Aspartame metabolism in animals. In: Aspartame Physiology and Biochemistry (Stegink LD, Filer LJ Jr, eds). New York: Dekker, 141-159.
- Piergorsh WW, Bailer AJ. 1997. Statistics for Environmental Biology and Toxicology. London: Chapman.
- Portier CJ, Bailer AJ. 1989. Testing for increased carcinogenicity using a survival-adjusted quantal response test. Fundam Appl Toxicol 12:731-737.
- Rall DP. 1995. Can laboratory animal carcinogenicity studies predict cancer in exposed children? Environ Health Perspect 103 suppl 6:173-175.
- Ranney RE, Opperman JA, Maldoon E, McMahon FG. 1976. Comparative metabolism of aspartame in experimental animals and humans. J Toxicol Environ Health 2:441-451.
- Soffritti M, Belpoggi F, Cevolani D, Guarino M, Padovani M, Maltoni C. 2002a. Results of long-term experimental studies on the carcinogenicity of methyl alcohol and ethyl alcohol in rats. In: Carcinogenesis Bioassays and Protecting Public Health.

Commemorating the Lifework of Cesare Maltoni and Collegues, 29-30 April 2002, New York, USA. Ann NY Acad Sci, 982:46-69.

- Soffritti M, Belpoggi F, Degli Esposti D, Lambertini L. 2005. Aspartame induces lymphomas and leukaemias in rats. Eur J Oncol 10:107-116.
- Soffritti M, Belpoggi F, Lambertini L, Lauriola M, Padovani M, Maltoni C. 2002b. Results of long-term experimental studies on the carcinogenicity of formaldehyde and acetaldhyde in rats. In: Carcinogenesis Bioassays and Protecting Public Health. Commemorating the Lifework of Cesare Maltoni and Collegues, 29-30 April 2002, New York, USA. Ann NY Acad Sci, 982: 87-105.
- Soffritti M, Belpoggi F, Minardi F, Bua L, Maltoni C. 1999. Mega-experiments to identify and assess diffuse carcinogenic risks. In: Uncertainty in the Risk Assessment of Environmental and Occupational Hazards, 24-26 September 1998, Bologna, Italy. Ann NY Acad Sci, 895:34-55.
- Soffritti M, Belpoggi F, Minardi F, Maltoni C. 2002c. Ramazzini Foundation cancer program: history and major projects, life-span carcinogenicity bioassay design, chemicals studied, and results. In: Carcinogenesis Bioassays and Protecting Public Health. Commemorating the Lifework of Cesare Maltoni and Collegues, 29-30 April 2002, New York, USA. Ann NY Acad Sci, 982:26-45.
- Soffritti M, Belpoggi F, Minardi F, Pinto C, Maltoni C. 1992. Chemopreventive effects of vitamin A (retinyl acetate and palmitate) and N-(4-hydroxyphenyl)retinamide in rats, with reference to mammary carcinoma. In: Progress and Perpectives in Chemoprevention of Cancer, March 1991, Milan, Italy. Raven Press, 79:47-60.

- Soffritti M, Belpoggi F, Padovani M, Lauriola M, Degli Esposti D, Minardi F. 2004. Lifetime carcinogenicity bioassay of toluene given by stomach tube to Sprague-Dawley rats. Eur J Oncol 9:91-102
- Soffritti M, Maltoni C, Maffei F, Biagi R. 1989. Formaldehyde: an experimental multipotent carcinogen. Toxicol Ind Health 5:699-730.
- Stegink LD. 1984. Aspartate and glutamate metabolism. In: Aspartame Physiology and Biochemistry (Stegink LD, Filer LJ Jr, eds). New York: Dekker, 47-76
- Tomatis L, Aitio A, Wilbourn J, Shuker L 1989. Human carcinogens so far identified. Jpn J Cancer Res 80:795-807.

No.	Products	No. of	Anima	Status of studies <sup>a</sup>		
		bioassays	Species	No.		
1.	Water in polyvinylchloride bottles	2	Rat <sup>b</sup>	2200	P <sup>c</sup>	
2.	Coca-cola	4	Rat <sup>b</sup>	1999	RP	
3.	Pepsi-Cola	1	Rat	400	Е	
4.	Ethyl alcohol (10% v/v)	4	Rat <sup>b</sup> , mice	1458	$\mathbf{P}^{\mathrm{d}}$	
5.	Sucrose	1	Rat	400	E	
6.	Aspartame	6	Rat, mice <sup>b</sup>	4460	BO, PP <sup>e</sup>	
7.	Sucralose	1	Mice <sup>b</sup>	760	BO	
8.	Caffeine	1	Rat	800	Е	
9.	Vitamin A	5	Rat	5100	$PP^{f}$	
10.	Vitamin C	5	Rat	3680	Е	
11.	Vitamin E	5	Rat	3680	Е	
12.	Feed sterilized by gamma rads	1	Rat <sup>b</sup>	2000	E	
TO	ΓAL	36		26937		

Table 1. Beverages and diet products studied at the CMCRC/ERF: status of studies

<sup>a</sup> P = published; PP = partially published; RP = ready for publication; E = in elaboration; BO = biophase ongoing <sup>b</sup> Treatment started from embryonal life; <sup>c</sup> Maltoni et al. 1997; <sup>d</sup> Soffritti et al. 2002a; <sup>e</sup> Soffritti et al. 2005;

<sup>f</sup> Soffritti et al. 1992

Dose ppm	Animals at start	imals Malignant tumors start				Tota bearing	al animals g lymphomas	Animals bearing dysplastic lesions and carcinomas of the renal pelvis and ureter <sup>a</sup>								1	Animals bearing peripheral nerve malignant Schwannomas					
(mg/kg b.w.)		Tumor-bearing animals		Total tumors		and leukemias		Dysplastic hyperplasias		Dysplastic papillomas		Carcinomas		Total		Cranial		Other sites		Total <sup>a,b</sup>		_
		No.	%	No.	Per 100 animals	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	_
100,000 (5,000)	100	43	43.0	55	55.0	29	29.0	3	3.0	0	_	1	1.0	4	4.0	3	3.0	1	1.0	4	4.0	_
50,000 (2,500)	100	38	38.0	45	45.0	20	20.0	2	2.0	0	-	1	1.0	3	3.0	3	3.0	0	-	3	3.0	
10,000 (500)	100	34	34.0	42	42.0	15	15.0	2	2.0	0	-	1	1.0	3	3.0	2	2.0	0	-	2	2.0	
2,000 (100)	150	60	40.0	69	46.0	33	22.0	4	2.7	0	-	1	0.7	5	3.3	2	1.3	0	-	2	1.3	
400 (20)	150	48	32.0	52	34.7	25	16.7	4 <sup>c</sup>	2.7	1 <sup>c</sup>	0.7	0 <sup>c</sup>	-	5 <sup>c</sup>	3.4	1	0.7	2	1.3	3	2.0	
80 (4)	150	44	29.3	49	32.7	23	15.3	3°	2.0	0 <sup>c</sup>	-	0°	-	3°	2.0	1	0.7	0	-	1	0.7	- 30
0 (0)	150	53	35.3*	59	39.3	31	20.7 **	1	0.7	0	-	0	-	1	0.7	1	0.7	0	-	1	0.7 *#	I

Table 2. Incidence of the preneoplastic and neoplastic lesions in male Sprague-Dawley rats in a life-span feed carcinogenicity study of Aspartame

<sup>a</sup> The tumor rates are based on the number of animals examined (necropsied)
<sup>b</sup> Near the control incidence are the p-values associated with the trend test

<sup>c</sup> Tissues from 149 animals have been analyzed

\* Statistically significant (p≤0.05) using Cochran-Armitage test

<sup>#</sup> Statistically significant (p≤0.05) using poly-k-test (k=3)

Dose ppm	Animals at start		Maligr	mors	Tota bearing	l animals lymphomas	Animals bearing dysplastic lesions and carcinomas of the renal pelvis and ureter <sup>a</sup>								Animals bearing peripheral nerve malignant Schwannomas							
(mg/kg b.w.)		Tumor-l anin	r-bearing imals		Total tumors	and leukemias		Dysplastic hyperplasias		Dysplastic papillomas		Carcinomas		Total		Cranial		Other sites		Total <sup>a,b</sup>		
		No.	%	No.	Per 100 animals	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	
100,000 (5,000)	100	51	51.0	64	64.0	25	25.0**	8	8.0	3	3.0	4	4.0#	15	15.0##	1	1.0	1	1.0	2	2.0	
50,000 (2,500)	100	58	58.0##	84	84.0	25	25.0##	6 <sup>e</sup>	6.1	1 <sup>e</sup>	1.0	3 <sup>e</sup>	3.0	10 <sup>e</sup>	10.1 ##	1	1.0	0	-	1	1.0	
10,000 (500)	100	40	40.0	62	62.0	19	19.0#	6	6.0	1	1.0	3(4)	3.0	10	10.0##	1	1.0	0	-	1	1.0	
2,000 (100)	150	67	44.7	86	57.3	28	18.7#	6	4.0	1	0.7	3(4)	2.0	10	6.7#	1	0.7	2	1.3	3	2.0	
400 (20)	150	70	46.7	95	63.3	30	20.0##	5	3.3	1	0.7	3	2.0	9	6.0#	0	_	0	-	0	_	
80 (4)	150	64	42.7	85	56.7	22	14.7	4	2.7	1	0.7	1	0.7	6	4.0	1	0.7	1	0.7	2	1.3	- 31
0 (0)	150	55	36.7 **	69	46.0	13	8.7 ***	2	1.3**	0	_ *	0	-	2	1.3 **##	0	-	0	-	0	_	I

Table 3. Incidence of the preneoplastic and neoplastic lesions in female Sprague-Dawley rats in a life-span feed carcinogenicity study of Aspartame

<sup>a</sup> The tumor rates are based on the number of animals examined (necropsied)

<sup>b</sup> Near the dosed group incidence are the p-values corresponding to pairwise comparisons between the controls and that dosed group

<sup>c</sup> Near the control incidence are the p-values associated with the trend test

<sup>d</sup> Between parentheses the numbers of tumors (one animal can bear bilateral tumors)

<sup>e</sup> Tissues from 99 animals have been analyzed

\* Statistically significant (p≤0.05) using Cochran-Armitage test

\*\* Statistically significant (p≤0.01) using Cochran-Armitage test

<sup>#</sup> Statistically significant ( $p \le 0.05$ ) using poly-k-test (k=3)

<sup>##</sup> Statistically significant (p≤0.01) using poly-k-test (k=3)

Dose	Animals	imals Lymphomas-Leukemias <sup>a</sup>																
ppm (mg/kg b.w.)	at start		Total <sup>b</sup>		Lymphoblastic lymphoma		Lymphoblastic leukemia		Lymphocytic lymphoma		vimmunoblastic mphoma	H	istiocytic arcoma	M le	onocytic zukemia	Myeloid leukemia		
		No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	
100,000 (5,000)	100	25	25.0 **	1	4.0	0	_	2	8.0	11	44.0	7	28.0	2	8.0	2	8.0	
50,000 (2,500)	100	25	25.0 ##	2	8.0	0	-	0	_	10	40.0	8	32.0	4	16.0	1	4.0	
10,000 (500)	100	19	19.0#	2	10.5	0	-	2	10.5	3	15.8	10	52.6	2	10.5	0	-	
2,000 (100)	150	28	18.7 *	5	17.8	1	3.6	1	3.6	8	28.6	8	28.6	4	14.3	1	3.6	
400 (20)	150	30 <sup>c</sup>	20.0 ##	7	23.3	0	_	2	6.7	8	26.7	9	30.0	5	16.7	0	-	
80 (4)	150	22	14.7	3	13.6	0	-	5	22.7	6	27.2	6	27.3	2	9.1	0	-	
0 (0)	150	13	8.7 ***	2	15.4	0	_	2	15.4	5	38.5	4	30.8	0	-	0	-	

Table 4. Incidence and distribution by hystocytotype of lymphomas-leukemias in female Sprague-Dawley rats in a life-span feed carcinogenicity study of Aspartame

<sup>a</sup> Percentages refer to the total number of animals bearing lymphomas-leukemias

<sup>b</sup> Percentages refer to the number of animals at start

<sup>c</sup> One animal bears two types of neoplasias: lymphoblastic lymphoma and histiocytic sarcoma

\*\* Statistically significant (p≤0.01) using Cochran-Armitage test

<sup>#</sup> Statistically significant (p≤0.05) using poly-k-test (k=3)

<sup>##</sup> Statistically significant (p≤0.01) using poly-k-test (k=3)





Figure 1. A. Mean daily feed consumption in males. B. Mean daily feed consumption in females. C. Mean body weights in males (M) and females (F). D. Survival in males. E. Survival in female.  $(-- \blacklozenge --$ 100,000 ppm;  $-- \blacktriangle -- 50,000$  ppm;  $-- \blacksquare -- 10,000$  ppm;  $-- \blacklozenge -- 2,000$ ppm;  $-- \between -- 400$  ppm;  $-- \between -- 80$ ppm;  $-- \between -- 80$  ppm;  $-- \between -- 80$ 



Figure 2. A. Carcinoma of the renal pelvis in a female rat administered 100,000 ppm aspartame in feed. Hematoxylin-Eosin (HE) X25. Scale bar 500 μm. B. A detail of the carcinoma shown in A. HE X400. Scale bar 20 μm. C. Malignant schwannoma of cranial nerves resembling Antoni B type pattern in a male rat administered 100,000 ppm aspartame in feed. HE X200. Scale bar 50 μm. D. Immunohistochemical characterization with S-100 protein of the schwannoma shown in C. X1000. Scale bar 10 μm.